

Western blotting

Supply and Reagents:

10XPBS (1L):

- 80 g NaCl
 - 2 g KCL
 - 14.4 g Na₂HPO₄ (dibasic, anhydrous)
 - 2.4 g KH₂PO₄ (monobasic anhydrous)
- Adjust pH to 7.4, autoclave.

PBST: 100ml

- 10X PBS 100ml
 - Tween 20 1ml
 - dd water 900ml
- Keep at room temperature

Transfer buffer (1L)

- Glycine 14.4 g
 - Tris 3.0g
 - Methanol 200ml
- Do not adjust pH by adding acid or bas, Keep @ RT

Other:

Thick filter paper (Bio-Rad)

Immobilon-P (10X15cm)

Supersignal West Pico Chemiluminescent Sbustrate (Pierce #34077)

X-ray file (X-OMAT)

Tap

Timer

Radioatography cassette

Protocol:

1. Isolate protein, run SDS-PAGE gel.

2. Blotting

1. Equilibrate the gel in transfer buffer for 10min
2. PVDF membrane was put in methanol for 1min, then put in transfer buffer
3. Sandwich the gel and membrane
(from bottom to top: filter paper, membrane, gel, filter paper; depends)
4. Transfer the protein at 0.2 A, 30min, Do not exceed 25V with this instrument.
5. Wash the membrane with PBST 20ml, 5 min
6. Mark the marker protein band by pencil (optional)

(The membrane can be dry for 30mins, treated with methanol, and PBST before use)

3. Blocking: in 20ml

Blocking solution	50ml
Nonfat dry milk	2.5g
PBST	50ml

Transfer the membrane to blocking solution.

Gently agitated for 1h @ RT

4. Reaction with antibody: in 10-15ml

Antibody solution:

Anti-His HPR antibody (Invitrogen)	1ul
PBST	10-15ml

(if use primary antibody, dilution could be 1: 10000 or higher)

Discard blocking solution

Wash lightly by PBST 20ml, 5min, twice, with agitating

Transfer the membrane to a tray containing antibody solution

Gently agitate overnight at cold room (12h)

***If you don't use secondary antibody, skip to Step 7.**

5. **Wash** membrane with 20ml PBST, 3 times, 10 min each
6. **Add secondary antibody** (1:10000~1:20000 dilution in 15-20 ml PBST), agitate for 60 min at room temperature

7. **Wash** membrane with 20ml PBST, 3 times, 10 min each.
8. **Chemiluminescence, exposure, and detection** at dark room

Hybridization bag

Radioautography cassette

X-Ray film

SuperSignal West Pico Chemiluminescent Substrate (Pierce #34077)

Tape

Timer

Parafilm

Micropipette

Kim-wipe

Turn on the light. Wear gloves.

Place the membrane on parafilm.

Prepare working solution (1:1 mixing) (0.5ml each)

Add onto the membrane

Incubate for 5 min

Remove excess liquid with an absorbent tissue

Place the membrane in hybridization bag

Put the membrane in cassette

Turn off light

Exposure X-ray film in cassette for 60s, 1min or 2min(depending on signal strength)

Develop the film.